

**ANTIFUNGAL ACTIVITY AND GENETIC DIVERSITY OF SELECTED  
*Pseudomonas* spp. FROM MAIZE RHIZOSPHERE IN VOJVODINA**

Dragana JOŠIĆ<sup>1</sup>, Radmila PIVIĆ<sup>1</sup>, Miroslav MILADINOVIĆ<sup>1</sup>, Mira STAROVIĆ<sup>2</sup>,  
Snežana PAVLOVIĆ<sup>3</sup>, Simonida ĐURIĆ<sup>4</sup>, Mirjana JARAK<sup>4</sup>

<sup>1</sup>Institute of Soil Science, Belgrade, Serbia

<sup>2</sup> Institute of Plant Protection and Environment, Belgrade, Serbia

<sup>3</sup> Institute for Medicinal Plant Research "Dr Josif Pancic", Belgrade, Serbia

<sup>4</sup>University of Novi Sad, Faculty of Agriculture, , Novi Sad, Serbia

Jošić D., R. Pivić, M. Miladinović, M. Starović, S. Pavlović, S. Đurić, and M. Jarak (2012): *Antifungal activity and genetic diversity of selected Pseudomonas spp. from maize rhizosphere in Vojvodina*. - Genetika, Vol 44, No. 2, 377 - 388.

Antibiotic production by plant-associated microorganisms represents an environmentally compatible method of disease control in agriculture. However, a wide application of bacterial strains needs careful selection and genetic characterization. In this investigation, selected *Pseudomonas* strains were characterized by rep-PCR methods using ERIC and (GTG)<sub>5</sub> primers, and partial 16S rDNA sequence analysis. None of strains produced

---

*Corresponding author:* Dragana Jošić, Institute of Soil Science, Genetic Laboratory, T. Drajzera 7, Belgrade, Serbia, E-mail: josicdragana@yahoo.com, Tel./Fax: 011/2667123, mob. 069/8223531; 063/8620622

homoserine lactones (C<sub>4</sub>, C<sub>6</sub>, C<sub>8</sub>) as quorum sensing signal molecules. Very poor production of phenazines and no significant fungal inhibition was observed for PS4 and PS6 strains. High amount of phenazines were produced by *Pseudomonas* sp. strain PS2, which inhibited mycelial growth of 10 phytopathogenic fungi in percent of 25 (*Verticillium* sp.) to 65 (*Fusarium equiseti*). Genetic characterization of the *Pseudomonas* sp. PS2 and evaluation of phenazines production, as the main trait for growth inhibition of phytopathogenic fungi, will allow its application as a biosafe PGPR for field experiments of plant disease control.

*Key words:* antifungal activity, ERIC, (GTG)<sub>5</sub>, PGPR, phenazines, *Pseudomonas*

#### INTRODUCTION

Plant growth-promoting bacteria (PGPB) are commonly present in many environments (COMPANT *et al.*, 2005). PGPB colonizing the root surfaces and the closely adhering soil interface - rhizosphere, are plant growth-promoting rhizobacteria (PGPR) (KLOEPPER *et al.*, 1999). Many PGPR strains produce secondary antimicrobial metabolites and can alter the environmental conditions for the growth of other microorganisms (NEWTON and FRAY, 2004). Among the secondary metabolites a variety of antibiotics have been identified, including phenazine, pyoluteorin, pyrrolnitrin, tensin, amphisin, 2,4-diacetylphloroglucinol (DAPG), oomycin A, tropolone, and cyclic lipopeptides produced by different pseudomonads (RAAIJMAKERS *et al.*, 2002; NIELSEN *et al.*, 2002; NIELSEN and SØRENSEN, 2003), macrolide antibiotic oligomycin A, produced by *Streptomyces* (KIM *et al.*, 1999) and kanosamine and zwittermicin A, produced by *Bacillus* (COMPANT *et al.*, 2005). A general hypothesis regarding these antibiotic metabolites is that they have a competitive function by inhibiting the growth of other microorganisms. Ability of PGPR to suppress fungal growth by production of some of these antibiotics was used in biological control of phytopathogens (KLOEPPER *et al.*, 1999; NIELSEN *et al.*, 2002; NIELSEN and SØRENSEN, 2003; WELLER, 2007). The mechanisms of biocontrol include competition for an ecological niche or a substrate, production of inhibitory chemicals, and induction of systemic resistance (ISR) in host plants to a broad spectrum of pathogens (HAAS *et al.*, 2000; BLOEMBERG and LUGTENBERG, 2001; GERHARDSON, 2002; HAAS *et al.*, 2002).

In cultivation of medicinal and aromatic plants (MAPs), antibiotic production by plant-associated microorganisms represents an environmentally acceptable method of disease control. Use of synthetic fungicides for disease suppression is limited due to a low efficiency, phytotoxicity, development of resistance, long degradation periods, environmental pollution and side effects on human and animal health. Phytopathogenic fungi are responsible for several plant diseases in different medicinal plants and cause very important economic losses in Serbian plantation (PAVLOVIC, 2008). Indigenous population of PGPR may prevent or suppress plant disease caused by phytopathogenic fungi. In earlier work, we isolated dominant pseudomonads and selected three plant growth-promoting *Pseudomonas* isolates from the maize rhizosphere in Vojvodina, Serbia. In attempt

to characterize them before application, we choose rep-PCR method for this investigation. Interspersed repetitive DNA sequence elements (rep) present in the prokaryote genomes are very useful as primers sites for genomic DNA amplification and yield complex genomic fingerprints suitable for estimation of genetic diversity. Enterobacterial Repetitive Intergenic Consensus (ERIC) sequence and BOX element are located in distinct, intergenic positions all around the chromosome (DE BRUIJN, 1992). The genetic differences between PGP *Pseudomonas* were already estimated using this method (BOTELHO, 2001; RANGARAJAN *et al.*, 2001; LANDA *et al.*, 2002; JOSIC *et al.*, 2012a).

The aim of this study was to investigate the genetic diversity of the selected soil bacterial strains using rep-PCR methods, estimate production of phenazines and to test their antimicrobial activity against fifteen pathogenic fungi originated from medicinal plants.

#### MATERIALS AND METHODS

**DNA manipulations.** Bacterial isolates were grown in liquid King B medium, at 28°C for 24h. Cell suspension was adjusted to  $A_{600nm} \approx 0.625$  by sterile distilled water. Total DNA was extracted after cells suspensions were boiled for 10 min, immediately cooled on ice, centrifuged and the supernatants were kept at -20°C or used for the PCR assay.

**rep-PCR.** Amplifications were performed in Eppendorf MasterCycler personal (Germany) with the temperature profile as reported by DE BRUIJN (1992). For the (GTG)<sub>5</sub> primer the cycles used were: an initial denaturation at 95°C for 7 min followed by 35 cycles of a three-step PCR program (94°C for 1 min, 52°C for 1 min and 65°C for 8 min) with a final extension at 65°C for 16 min. ERIC1R/2 primer set were used for additional fingerprinting under following amplification conditions: initial denaturation at 95°C for 3 min, 35 cycles of 94°C for 1 min, 53°C for 1 min, and 65°C for 8 min with the final extension at 65°C for 16 min. PCR products were visualised by electrophoresis on 1.5% (w/v) agarose gels in 0,5 Tris-borate-EDTA buffer for 8h at 3 V/cm, stained with ethidium bromide and photographed under UV transillumination. BOX and ERIC PCR fingerprint results were converted to binary form and cluster analysis of strains were done with the program STATISTICA 7.

**PCR amplification of bacterial 16S rRNA gene.** The 16S rRNA gene was amplified by using universal bacterial primers fD1 and rD1 (WEISBURG *et al.*, 1991). PCR amplification was carried out in a 50 µl reaction volume using Green Taq Dream master mix (Fermentas, Lithuania) with 1µl of template DNA and 0.1 µmol of each primers fD1 and rD1. DNA amplification was performed with the following temperature profile: an initial denaturation at 95°C for 4 min, followed by 35 cycles of denaturation (1 min at 94°C), annealing (1 min at 58°C), and extension (2 min at 72°C); with a final extension at 72°C for 5 min. Amplified DNA was examined by electrophoresis for 2h at 5V/cm in 1,2% (w/v) agarose gel with GeneRuler DNA Ladder mix SM0331 (Fermentas, Lithuania).

To determine the species identity, sequences of bacterial isolates were compared with the published sequences on the NCBI GenBank database using the algorithm

BLAST. The 16S rDNA sequences of bacterial strains are deposited under accession numbers JQ818423, JQ818424 and JQ818424.

**Production of phenazines and quorum sensing signal molecules.**

Overnight culture of bacterial isolates grown in King B liquid medium were subjected for phenazine and 2-OH-phenazine extraction following the method of CHANCEY *et al.* (1999). Quantification of phenazine and 2-OH-phenazine were performed by the UV light spectroscopy measuring absorbance at 367 nm and 468 nm, respectively (MADDULA *et al.* 2008).

Production of N-acyl-homoserinelactone (AHLs) as quorum sensing (QS) signal molecules by maize-rhizosphere isolates was tested by using a AHLs-sensor strain *Chromobacterium violaceum*, as described by PORITSANOS *et al.* (2006). Briefly, bacterial isolate and *Chromobacterium violaceum* were inoculated on LB medium as an opposite lines at distance of 3 mm, and incubated for four days at 25°C. A purple pigment production by *Chromobacterium violaceum* indicated on no long chain AHLs production.

***In vitro* antifungal activity.** The bacterial isolates were screened for their ability to inhibit growth of 15 medical plant pathogenic fungi: *Alternaria alternata*, *Dreschlera tetramera*, *Curvularia lunata*, *Diaporthe eres complex*, *Fusarium semitectum*, *Fusarium sporotrichioides*, *Fusarium equiseti* from *Salvia officinalis* L., *Fusarium equiseti* from *Matricaria chamomilla* L., *Myrothecium verrucaria*, *Verticillium* sp., *Fusarium oxysporum*, *Fusarium verticillioides*, *Fusarium proliferatum*, *Fusarium solani*, and *Sclerotinia sclerotiorum* (PAVLOVIC, 2008). The growth of a fungal colony (6 mm plugs placed in the middle of plate with Waksman agar) was determined after 7 days at 25°C, depending on the growth speed of the fungus. Control treatments were the fungal inoculums used alone, since the bacterial inoculation were placed 30 mm from fungal inoculum. The assay was performed four times for each bacterial isolates. The mycelium length was measured towards and away from bacterial inoculum. The percentage of fungal growth inhibition was calculated as described by ZARRIN *et al.* (2009): % Inhibition = [(R-r)/R] x 100, where r is the length of the fungal colony opposite the bacterial colony and R is the maximum radius of the fungal colony away from the bacterial colony.

## RESULTS

**rep-PCR based genotyping and 16S rRNA gene amplification.** Genomic fingerprints were obtained by amplification with BOX or enterobacterial repetitive intergenic consensus sequence (ERIC) primers. BOX fingerprints by (GTG)<sub>5</sub> primer were used as a first step of genodiversity observation (Figure 1a). ERIC-PCR confirmed differences in repetitive elements dispersion (Figure 1b). rep-PCR amplifications of the strains yielded complex genomic fingerprints consisting amplicons ranging from 100 to 3000 bp in size. Cluster analysis of genomic fingerprint patterns separate PS6 strain from PS2 and PS4 and showed 67.8 % of diversity. Strains PS2 and PS4 showed 59% patterns dissimilarity (Figure 2).

16S rDNA gene sequences of 1500 bp were obtained from each strain by PCR amplification with the conserved eubacterial primers fD1 and rD1 defined by

WEISBURG *et al.* (1991). Partial sequencing of the PCR products from the selected strains confirmed high level of genetic diversity.

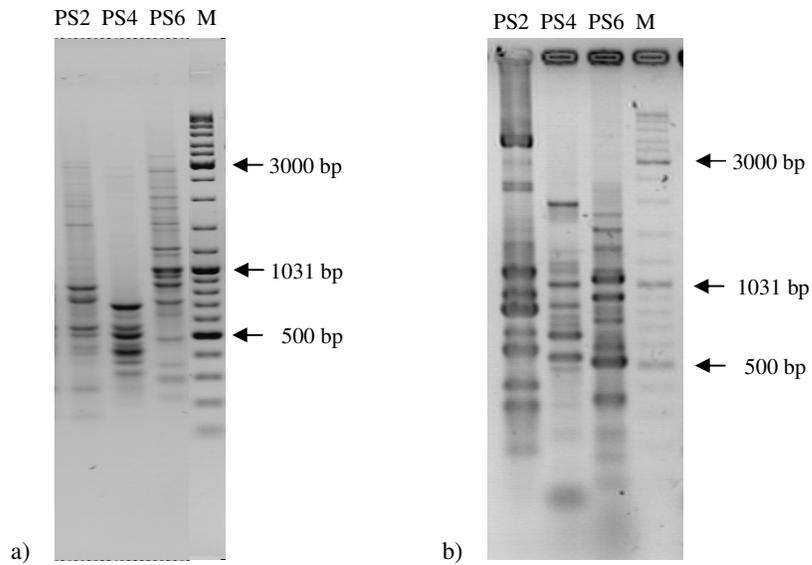


Figure 1. Genomic DNA fingerprint patterns generated by a) (GTG)<sub>5</sub> and b) ERIC PCR amplifications

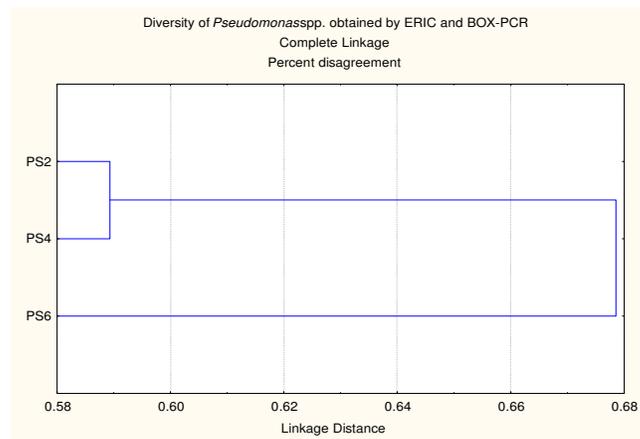


Figure 2. Cluster analysis of genomic fingerprint patterns generated by (GTG)<sub>5</sub> and ERIC PCR amplifications of whole-cell genomic DNA

**Production of phenazines and quorum sensing signal molecules.** The production of phenazines was evaluated by measuring absorbance of phenazine-carboxylic acid (PCA) at 367 nm and 2-hydroxi-phenazine-carboxylic acid (2-OH-PCA) at 468 nm. Although the three strains were producing both antibiotics, 2-OH-PCA was produced predominantly from each of them. The Strain PS2 synthesized significantly more amount phenazines (both PCA and 2-OH-PCA) (Table 1). *In vitro* production of the signal molecules was evaluated in a plate with *Chromobacterium violaceum* CV026 as an indicator strain. The results showed that none of the strains was able to produce exogenous C<sub>4</sub>, C<sub>6</sub> or C<sub>8</sub> homoserine lactones as quorum sensing signal molecules.

Table 1. Production of acyl-homoserinelactones and phenazines by *Pseudomonas* spp. strains from maize rhizosphere

Strain	AHLs	Absorbance of	
		PCA <sup>a</sup>	2-OH-PCA <sup>b</sup>
PS2	-	0.160 ± 0.016 B <sup>c</sup>	0.194 ± 0.010 B
PS4	-	0.019 ± 0.002 A	0.042 ± 0.006 A
PS6	-	0.007 ± 0.002 A	0.029 ± 0.004 A

<sup>a</sup> Determination of phenazine-carboxylic acid (PCA) production at absorbance at 367 nm.

<sup>b</sup> Determination of production of 2-hydroxi-phenazine-carboxylic acid (2-OH-PCA) at absorbance at 468 nm

<sup>c</sup> values in the columns marked with the same letter are not statistically significant according to the Duncan's test (P=0,05)

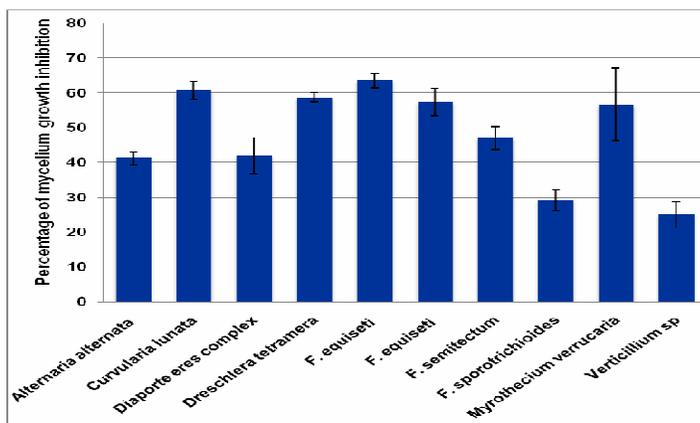


Figure 3. *In vitro* bioassay of fluorescent *Pseudomonas* strain PS2 against phytopathogenic fungi isolated from medical plants

***In vitro* antifungal activity.** For a further characterization of a selected strain PS2 with potential for plant growth promotion, dual culture were performed using fifteen phytopathogenic fungi known for their detrimental effects on medical plants. None of the bacteria tested improved growth of the phytopathogenic fungi. *Pseudomonas* PS2 strain was induced hifal deformation and growth suppression at margin with bacteria on *Fusarium oxysporum*, *F. verticillioides*, *F. proliferatum*, *F. solani*, and *Sclerotinia sclerotiorum*. The valuable percentage of mycelia growth inhibition of ten phytopathogenic fungi were observed only for PS2 strain (Figure 3). *Pseudomonas* PS4 strain caused hyphal deformation of *Diaporthe eres complex* near to bacterial colonies. Growth of *Dreschlera tetramera* and *F. equiseti* were inhibited by PS6 strain less then 10%.

#### DISCUSSION

Three distinct genotypes of investigated *Pseudomonas* strains were differentiated by amplification of ERIC and BOX elements. Strains PS2 and PS4 clustered separately from PS6 strain, but all strains showed very high dissimilarity level (59-67.8 %). Differences in phenotypic and plant growth promoting properties of investigated strains in previous investigation (DJURIC *et al.*, 2011) are in agreement with high genetic diversity obtained in this work. *Pseudomonad* strains from soybean rhizosphere were analysed by BOX and ERIC - PCR (BOTELHO, 2001) and similar profiles were observed to the *Pseudomonas fluorescens* strain 2-79 which is phenazine-producer. To assess the genotypic diversity of *Pseudomonas* collection from wheat rhizosphere, MCSPADEN GARDENER *et al.* (2000) compared their ERIC and BOX-PCR patterns to 4 previously described 2,4-DAPG producers. LANDA *et al.* (2002) reported the relationship between pea rhizosphere competence and genotype. Genetic profiles generated by BOX-PCR correlate very well with RFLP profiles of **phlD** genes and this relationship will allow a more directed selection of 2,4-DAPG-producing strains for use in biological control.

Fluorescent *Pseudomonas* has been suggested as potential biological control agent (HAS and DEFAGO, 2005) due to its ability to colonize rhizosphere and protect plants against a wide range of important agronomic fungal diseases such as damping-off of sugar beet (KUMAR *et al.*, 2002) and damping-off of cotton with *Rhizoctonia solani* as a causal agent (PAL *et al.*, 2000). Fluorescent pseudomonad antagonizes all the reproductive phases of the *Phytophthora capsici*, the causal organism of foot rot disease (PAUL and SARMA, 2006) and exhibits strong antifungal activity against *Rhizoctonia solani* (REDDY *et al.*, 2008), *R. bataticola* and *Fusarium oxysporum* found in rice and sugarcane rhizosphere (KUMAR *et al.*, 2002). *Pseudomonas* shows biocontrol potential against phytopathogenic fungi *in vivo* and *in vitro* conditions from chickpea rhizosphere (SARAF *et al.*, 2008). Fluorescent *Pseudomonas* suppresses plants fungal diseases mainly through the production of antifungal metabolites. We detected lytic enzymes and HCN production in addition to siderophores and IAA production in our previous investigation of different fluorescent *Pseudomonas* from maize rhizosphere (DJURIC *et al.*, 2011). In this study

we investigated selected strains (PS2, PS4 and PS6) for short chain ACHLs and phenazines production. None of the strains produced a short chain ACHLs, two strains were characterized as very poor producers of 2-hydroxyl-phenazinecarboxylic acid and 1-phenazinecarboxylic acid, while strain PS2 produced high amount of both antibiotics. This strain produced higher amount of 2-OH-PCA than PCA. A greater antibiotic activity of 2-OH-PCA has been demonstrated against array of bacterial and fungal organisms in comparison to PCA (MADDULA *et al.*, 2008). PICARD *et al.* (2000) demonstrated that, in the field condition, maize is more sensitive to rhizosphere pathogens during the first stage of growth, which corresponds to the end of germination. They detected low density occurrence of an antibiotic DAPG (2,4-diacetylphloroglucinol) -producing population of *Pseudomonas* isolated from the maize rhizosphere during the first stage of maize growth, and highlighted the importance for releasing an effective bio-control agent during the first stage of maize development.

*Pseudomonas* PS2 showed effective mycelial growth inhibition of 10 out of 15 phytopathogenic fungi. The most significant changes in the mycelial growth rates was achieved against *Fusarium equisetii* isolated from sage (*Salvia officinalis*) and *Curvularia lunata*, inducing the growth suppression of 64% and 61%, respectively. The least inhibitory effect was detected on the mycelium extension of *Fusarium sporotrichoides* (25%). Appreciable efficacy of *P. fluorescens* against *Alternaria tenuissima* (~83%) and *F. solani* (~79%) reported SEETHA RAMULU *et al.* (2010). Biocontrol of *Alternaria tenuissima* originated from *Ocimum basilicum* L. using indigenous *Pseudomonas* spp. were reported recently (JOŠIĆ *et al.*, 2012b), showing that PS2 reduced 90% of disease incidence in greenhouse experiments with non-sterile soil. About 80% of the growth inhibition of *F. moniliformae* and *Colletotrichum acutatum* cause six days old culture filtrate of *P. fluorescens* MA-4 strain (MISHRA *et al.*, 2011). As previously shown, the phenazines are redox-active antibiotics (DIETRIX *et al.* 2008) and may contribute to iron mobilization in soils in addition to siderophores (HERNANDEZ *et al.*, 2004; WANG and NEWMAN, 2008).

#### CONCLUSION

Selected *Pseudomonas* strains isolated from maize rhizosphere showed high genetic diversity on the basis of ERIC and (GTG)<sub>5</sub> PCRs, which is confirmed by 16S rDNA sequences analysis. Ability to produce IAA and siderophores, as detected earlier, and substantial production of phenazines are main characteristic of *Pseudomonas* PS2 strain. Growth inhibition greater than 50% was observed for *Curvularia lunata*, *Myrothecium verrucaria*, *Dreschlera tetramera* and *Fusarium equisetii* isolated from two medicinal plants -*Salvia officinalis* L. and *Matricaria chamomilla* L. Further field trials will test the competitiveness of PS2 strain with indigenous population, effects in biological control of phytopathogenic fungi and possible advantage based on detected PGP traits.

## ACKNOWLEDGEMENTS

We are grateful to dr Milan Kojic for providing us with the indicator strain *Chromobacterium violaceum* used in this investigation. This study was supported by the Ministry of Education and Science of Republic of Serbia, Project No III 46007 (New indigenous bacterial isolates *Lysobacter* and *Pseudomonas* as an important sources of metabolites useful for biotechnology, plant growth stimulation and disease control: from isolates to inoculants).

Received December 2<sup>th</sup>, 2011

Accepted May 23<sup>rd</sup>, 2012

## REFERENCES

- COMPANT, S., B. DUFFY, J. NOWAK, C. CLEMENT and E. A. BARKA (2005): Use of Plant Growth-Promoting Bacteria for Biocontrol of Plant Diseases: Principles, Mechanisms of Action, and Future Prospects. *Appl. Environ. Microbiol.*, Volumen 71(9), pp. 4951–4959.
- BLOEMBERG, G. V. and B. J. J. LUGTENBERG (2001): Molecular basis of plant growth promotion and biocontrol by rhizobacteria. *Curr. Opin. Plant Biol.* Volumen 4, pp. 343–350.
- BOTELHO, G.R. (2001): Seleção de *Pseudomonas* fluorescentes para Controle Biológico da Podridão Vermelha da Raiz causada por *Fusarium solani*. Rio de Janeiro, pp. 108 (Dr. Thesis, Biotecnologia Vegetal, UFRJ).
- CHANCEY, S.T., D.W. WOOD and L.S. PIERSON III (1999): ***Two-component transcriptional regulation of n-acyl-homoserine lactone production in Pseudomonas aureofaciens.*** *Appl. Environ. Microbiol.* Volumen 65, pp. 2294 - 2299.
- DE BRUIJN F.J. (1992): Use of repetitive (repetitive extragenic palindromic and enterobacterial repetitive intergeneric consensus) sequences and the polymerase chain reaction to fingerprint the genomes of *Rhizobium meliloti* isolates and other soil bacteria. *Appl. Environ. Microbiol.* Volumen 58, pp. 2180-2187.
- DIETRICH, L.E.P., T.K. TEAL, A. PRICE-WHELAN and D.K. NEWMAN (2008): Redox-active antibiotics control gene expression and community behavior in divergent bacteria. *Science.* Volumen 321(5893), pp. 1203–1206.
- DJURIC, S., A. PAVIC, M. JARAK, S. PAVLOVIC, M. STAROVIC, R. PIVIC and D. JOSIC (2011): Selection of indigenous fluorescent pseudomonad isolates from maize rhizospheric soil in Vojvodina as possible PGPR. ***Rom Biotechnol Lett.*** Volumen 16, pp. 6580-6590.
- GERHARDSON, B. (2002): Biological substitutes for pesticides. *Trends Biotechnol.* Volumen 20, pp.338–343.
- HAAS, D., C. BLUMER and C. KEEL (2000): Biocontrol ability of fluorescent pseudomonads genetically dissected: importance of positive feedback regulation. *Curr. Opin. Biotechnol.* Volumen 11, pp. 290–297.
- HAAS, D., C. KEEL and C. REIMMANN (2002): Signal transduction in plant beneficial rhizobacteria with biocontrol properties. *Antonie Leeuwenhoek* Volumen 81, pp. 385–395.

- HAAS, D. and G. DÉFAGO (2005): Biological control of soil-borne pathogens by fluorescent pseudomonads. *Nat. Rev. Microbiol.* Volumen 3, pp. 307-319.
- HERNANDEZ, M.E., A. KAPPLER and D.K. NEWMAN (2004). *Phenazines and other redox-active antibiotics promote microbial mineral reduction.* *Appl. Envir. Microbiol.* Volumen 70, pp. 921 - 928.
- JOSIC D., DELIC D., RASULIC N., STAJKOVIC O., KUZMANOVIC D., STANOJKOVIC A., R. PIVIC (2012a). Indigenous Pseudomonads from Rhizosphere of Maize grown on Pseudogley Soil in Serbia. *Bulgarian Journal of Agricultural Science.* Volumen 18 (2), pp. 197-206.
- JOŠIĆ, D, S. PAVLOVIĆ, M. STAROVIĆ, S. STOJANOVIĆ, A. STANOJKOVIĆ-SEBIĆ, R. PIVIĆ (2012b). Biocontrol of *Alternaria tenuissima* originated from *Ocimum basilicum* L using indigenous *Pseudomonas* spp. Strains. 7<sup>th</sup> CMAPSEEC, Subotica, Serbia, 27<sup>th</sup>-31<sup>th</sup> May, 2012. Proceedings, pp. 195-200.
- KIM, B. S., S. S. MOON and B. K. HWANG (1999): Isolation, identification and antifungal activity of a macrolide antibiotic, oligomycin A, produced by *Streptomyces libani*. *Can. J. Bot.* Volumen 77, pp. 850–858.
- KLOPPER, J.W., R. RODRIGUEZ-UBANA, G.W. ZEHNDER, J.F. MURPHY, E. SIKORA, and C. FERNANDEZ (1999): Plant root-bacterial interactions in biological control of soilborne diseases and potential extension to systemic and foliar diseases. *Austral. Plant Pathol.* Volumen 28, pp. 21–26.
- KUMAR, N.R., V.T. ARASU and P. GUNASEKARAN (2002): Genotyping of antifungal compounds producing plant growth-promoting rhizobacteria, *Pseudomonas fluorescens*. *Current Science*, Volumen 82 (Suppl 12), pp. 1465-1466.
- LANDA, B.B., O.V. MAVRODI, J.M. RAAIJMAKERS, B.B. MCSPADEN GARDENER, L. S. THOMASHOW and D.M. WELLER (2002): Differential ability of genotypes of 2,4-diacetylphloroglucinol-producing *Pseudomonas fluorescens* strains to colonize the roots of pea plants. *Appl. Environ. Microbiol.* Volumen 68, pp. 3226–3237.
- MADDALA, V.S.R.K., E.A. PIERSON and L.S. PIERSON III (2008): Altering the ratio of phenazines in *Pseudomonas chlororaphis* (aureofaciens) strain 30-84: effects on biofilm formation and pathogen inhibition. *J. Bacteriol.* Volumen 190, pp. 2759 – 2766.
- MCSPADEN GARDENER, B.B., K.L. SCHROEDER, S.E. KALLOGER, J.M. RAAIJMAKERS, L.S. THOMASHOW and D.M. WELLER (2000): Genotypic and phenotypic diversity of *phlD*-containing *Pseudomonas* strains isolated from the rhizosphere of wheat. *Appl. Environ. Microbiol.* Volumen 66, pp. 1939–1946.
- MISHRA, R.K., O. PRAKASH, A.K. TIWARI, A. PANDEY, M. ALAM, and A. DIKSHIT (2011): Culture filtrate antibiosis of plant growth promoting rhizobacteria PGPRs against phytopathogens infecting medicinal and aromatic plants. *International Journal of Research in Biological Sciences.* Volumen 1, pp. 45–51.
- NEWTON, J.A., and R.G. FRAY (2004): Integration of environmental and host-derived signals with quorum sensing during plant-microbe interactions. *Cell. Microbiol.* Volumen 6, pp. 213–224.
- NIELSEN, T. H., D. SØRENSEN, C. TOBIASEN, J.B. ANDERSEN, C. CHRISTEOPHERSEN, M. GIVSKOV and J. SØRENSEN (2002): Antibiotic and biosurfactant properties of cyclic lipopeptides produced by fluorescent *Pseudomonas* spp. from the sugar beet rhizosphere. *Appl. Environ. Microbiol.* Volumen 68, pp. 3416–3423.
- NIELSEN, T. H. and J. SØRENSEN (2003): Production of cyclic lipopeptides by *Pseudomonas fluorescens* strains in bulk soil and in the sugar beet rhizosphere. *Appl. Environ. Microbiol.* Volumen 69, pp. 861–868.

- PAL, K.K., K.V.B.R. TILAK, A.K. SAXENA, R. DEY and C.S. SINGH (2000): Antifungal characteristics of a fluorescent *Pseudomonas* strain involved in the biological control of *Rhizoctonia solani*. Research in Microbiology, Volumen 155 (Suppl 3), pp. 233-242.
- PAUL, D. and Y.R. SARMA (2006): Antagonistic effects of metabolites of *Pseudomonas fluorescens* strains on the different growth phases of *Phytophthora capsici*, foot rot pathogen of black pepper (*Piper nigrum* L). Archives of Phytopathology and Plant Protection, Volumen 39 (Suppl 2), pp. 113-118.
- PAVLOVIC, S. (2009): Mycoses of more important medicinal plants in Serbia. PhD thesis. Faculty of Agriculture, University in Novi Sad.
- PICARD, C., F. DI CELLO, M. VENTURA, R. FANI and A. GUCKERT (2000): Frequency and Biodiversity of 2,4-Diacetylphloroglucinol-Producing Bacteria Isolated from the Maize Rhizosphere at Different Stages of Plant Growth Appl. Envir. Microbiol. Volumen 66, pp. 948 - 955.
- PORITSANOS, N., C. SELIN, W.G.D. FERNANDO, S. NAKKEERAN and T.R. KIEVIT (2006): A GacS deficiency does not affect *Pseudomonas chlororaphis* PA23 fitness when growing on canola, in aged batch culture or as a biofilm. Can. J. Microbiol. Volumen 52, pp. 1177-1188.
- RAAJMAKERS, J.M., M. VLAMI and J.T. DE SOUZA (2002): Antibiotic production by bacterial biocontrol agents. Antonie Leeuwenhoek Volumen 81, pp. 537-547.
- RANGARAJAN, S., P. LOGANATHAN, L.M. SALEENA and S. NAIR (2001): Diversity of pseudomonads isolated from three different plant rhizospheres. Journal of Applied Microbiology, Volumen 91 (Suppl 4), pp. 742-749.
- REDDY, B.P., K.R.N. REDDY, S.M. RAO AND K.S. RAO (2008): Efficacy of antimicrobial metabolites of *Pseudomonas fluorescens* against rice fungal pathogens. Current Trends in Biotechnology and Pharmacy, Volumen 2 (Suppl 1), pp. 178-182.
- SARAF, M., A. THAKKER and B.V. PATEL (2008): Biocontrol activity of different species of *Pseudomonas* against phytopathogenic Fungi *In vivo* and *In vitro* conditions. International Journal of Biotechnology & Biochemistry, Volumen 4 (Suppl 3&4).
- SEETHA RAMULU, J., G.R.C. RAJA and R. RAMANJANEYULU (2010): Evaluation of certain plant extracts and antagonists against *Fusarium solani* and *Alternaria tenuissima*, the incitants of root rot and die-back diseases of mulberry. Int. J. Indust. Entomol. Volumen 20, pp. 1-5.
- WANG, Y. and D.K. NEWMAN (2008): Redox reactions of phenazine antibiotics with ferric (hydr)oxides and molecular oxygen. Environ. Sci. Technol. Volumen 42, pp. 2380-2386.
- WEISBURG, W.G., S.M. BARNES, D.A. PELLETIER and D.J. LANE (1991): 16S ribosomal DNA amplification for phylogenetic study. J Bacteriol. Volumen 173, pp. 697-703.
- WELLER, D.M. (2007): *Pseudomonas* biocontrol agents of soilborne pathogens: looking back over 30 years. Phytopathology Volumen 97, pp. 250-256.
- ZARRIN, F., M. SALEEMI, M. ZIA, T. SULTAN, M. ASLAM, RIAZ -UR- REHMAN and M.F. CHAUDHARY (2009): Antifungal activity of plant growth-promoting rhizobacteria isolates against *Rhizoctonia solani* in wheat. Afr. J. Biotechnol. Volumen 8 (2), pp. 219-225

**ANTIFUNGALNA AKTIVNOST I GENETIČKI DIVERZITET  
SELEKTOVANIH *Pseudomonas* spp. IZ RIZOSFERE KUKURUZA  
U VOJVODINI**

Dragana JOŠIĆ<sup>1</sup>, Radmila PIVIĆ<sup>1</sup>, Miroslav MILADINOVIĆ<sup>1</sup>, Mira STAROVIĆ<sup>2</sup>,  
Snežana PAVLOVIĆ<sup>3</sup>, Simonida ĐURIĆ<sup>4</sup>, Mirjana JARAK<sup>4</sup>

<sup>1</sup>Institut za zemljište, Beograd, Srbija

<sup>2</sup>Institut za zaštitu bilja, Beograd, Srbija

<sup>3</sup>Institut za Medicinska istraživanja "Dr Josif Pancic", Beograd, Srbija

<sup>4</sup>Univerzitet Novi Sad, Poljoprivredni fakultet, Novi Sad, Serbia

Ekološki prihvatljiv metod kontrole bolesti bilja u poljoprivredi obuhvata primenu rizosfernih mikroorganizama koji produkuju antibiotike. Široka primena bakterija zahteva njihovu pažljivu selekciju i genetičku karakterizaciju. Ova istraživanja obuhvatila su karakterizaciju selektovanih *Pseudomonas* sojeva na osnovu rep-PCR metode korišćenjem ERIC i (GTG)<sub>5</sub> prajmera, kao i parcijalne sekvence 16S rDNA. Nije detektovana produkcija acil-homoserin laktona (C<sub>4</sub>, C<sub>6</sub>, C<sub>8</sub>) kao QS molekula. Sojevi PS4 i PS6 su vrlo oskudno produkovali fenazine i nisu značajno inhibirali rast fitopatogenih gljiva. Soj *Pseudomonas* sp. PS2 je obilno produkovao 2-OH-PCA i PCA i inhibirao rast micelija 10 fitopatogenih gljiva od 25% (*Verticillium* sp.) do 65% (*Fusarium equiseti*). Genetička karakterizacija soja *Pseudomonas* sp. PS2 i detekcija produkcije fenazina kao glavnog inhibitora rasta fitopatogenih gljiva omogućuje primenu ovog soja kao bezbednog PGP agensa za biološku kontrolu u budućim ogledima u polju.

Primljeno 02. XII. 2011.

Odobreno 23. V. 2012.